



Taxezipidine A, a Novel Taxoid from Seeds of Japanese Yew *Taxus cuspidata*

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Abstract: A novel taxoid, taxezipidine A (**1**), has been isolated from seeds of Japanese yew *Taxus cuspidata* Sieb. et Zucc. and the structure elucidated on the basis of spectroscopic data. Taxezipidine A (**1**) is the first taxoid with a hemiketal ring at C-11 ~ C-13, C-15, and C-17. © 1997 Elsevier Science Ltd.

In our continuing search for bioactive taxoids, we isolated previously new taxane and related diterpenoids containing various skeletons, taxuspines A ~ H and J ~ Z¹, from stems and leaves of Japanese yew *Taxus cuspidata* Sieb. et Zucc. Further investigation on extracts of seeds of this yew led to isolation of a novel taxoid, named taxezipidine A (**1**). In this paper we describe the isolation and structure elucidation of **1**.

The methanolic extract of seeds of the yew collected at Sapporo was partitioned between toluene and water and then the aqueous layer was extracted with chloroform. The chloroform-soluble portion was subjected to a silica gel column followed by reversed-phase column to afford taxezipidine A (**1**, 0.00032%).

Taxezipidine A (**1**)² was obtained as a colorless amorphous solid and the molecular formula was established to be C₂₆H₃₆O₉ by HRFABMS [*m/z* 493.2428 (M+H)⁺, Δ -1.0 mmu]. IR absorptions implied that **1** possessed hydroxy (3450 cm⁻¹) and ester (1740 cm⁻¹) groups. Analyses of the ¹H and ¹³C NMR data and HMQC spectrum of **1** provided three acetyls, one tetrasubstituted olefin, one exomethylene, one ketal carbon, four oxymethines, two methines, one oxymethylene, three methylenes, two quaternary

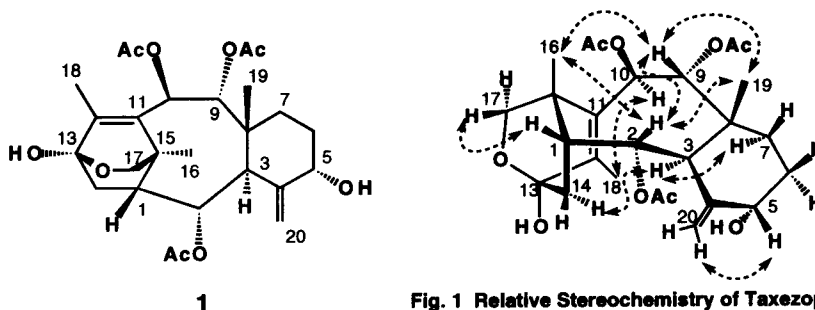


Fig. 1 Relative Stereochemistry of Taxezipidine A (**1**)
Dotted arrows denote NOESY correlation

carbons, and three methyl groups. Connectivities of C-1 to C-3, C-5 to C-7, C-9 to C-10, and C-14 to C-1 were deduced from the ^1H - ^1H COSY spectrum. In the decoupled HMBC (D-HMBC)³ spectrum, long-range ^1H - ^{13}C correlations of H-14a, H-17a, and H₃-18 to C-13 (δ 97.50), H-17a to C-1, C-11, and C-15, and H₃-18 to C-11 and C-12 indicated that **1** possessed an oxabicyclo[2.2.2]octene moiety consisting of C-11 ~ C-14, C-1, C-15, C-17, and O-17. HMBC correlations of H₃-18 to C-11, C-12, and C-13 and H₃-16 to C-1, C-11, and C-15 revealed that Me-18 and Me-16 were attached at C-12 and C-15, respectively. HMBC cross-peaks of H-3 to C-8, H-9 to C-8 and C-11, H-10 to C-15, H-1 to C-11 revealed the presence of an eight-membered ring, while the correlations of H-3 to C-4, H-20a to C-3 and C-5, and H-7 to C-3 implied the presence of a cyclohexane moiety with an exomethylene at C-4. Three acetoxy groups were attached at C-2, C-9, and C-10 based on HMBC correlations, while one of two hydroxy groups was connected to C-5 (δ_{H} 4.22). The remaining hydroxy group was connected to C-13 from comparison of ^{13}C NMR chemical shift of C-13 (δ_{C} 97.50) with those of hemiketal carbons (δ 96 ~ 98).⁴ Thus the structure of taxezopidine A was assigned to be **1**. Relative stereochemistry of **1** was deduced from NOESY data (Fig. 1) and ^1H - ^1H coupling constants. A boat-chair conformation of the eight-membered ring was elucidated from the coupling constant (10.7 Hz) between H-9 and H-10 and NOESY correlations, while a chair conformation of the cyclohexane ring was assigned from NOESY correlations. NOESY correlation between H-1 and H-17a indicated that both protons were quasi-axials and the oxymethylene (C-17) was β -oriented at C-15, while the correlations of H-3 to H-14a and H₃-18 revealed that **1** has a cage-like backbone conformation.

Taxezopidine A (**1**) is the first taxoid with a hemiketal ring at C-11 ~ C-13, C-15, and C-17 from yew trees. It is noted that **1** has a cage-like backbone conformation similar to usual taxoids consisting 6/8/6-membered ring system, although **1** contains the oxabicyclo[2.2.2]octene moiety.

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- Taxezopidine A (**1**): $[\alpha]_{\text{D}}^{27} +5.0^\circ$ (c 0.10, CHCl_3); IR (film) ν_{max} 3450, 1740, 1370, and 1020 cm^{-1} ; ^1H NMR (CDCl_3) δ 0.90 (3H, s, H-19), 1.54 (3H, s, H-16), 1.63 (1H, d, $J = 14.6$ Hz, H-14b), 1.66 (2H, m, H-6b and H-7b), 1.80 (1H, m, H-6a), 1.94 (1H, m, H-7a), 1.98 (3H, s, 10-AcO), 2.02 (1H, dd, $J = 14.6$ and 9.2 Hz, H-14a), 2.06 (3H, s, 9-AcO), 2.07 (3H, s, 2-AcO), 2.11 (1H, dd, $J = 9.2$ and 0.7 Hz, H-1), 2.20 (3H, s, H-18), 3.11 (1H, d, $J = 8.0$ Hz, H-17b), 3.26 (1H, d, $J = 5.3$ Hz, H-3), 3.49 (1H, d, $J = 8.0$ Hz, H-17a), 4.22 (1H, brs, H-5), 4.91 (1H, s, H-20b), 5.16 (1H, s, H-20a), 5.48 (1H, dd, $J = 5.3$ and 0.7 Hz, H-2), 5.71 (1H, d, $J = 10.7$ Hz, H-9), and 6.14 (1H, d, $J = 10.7$ Hz, H-10), ^{13}C NMR (CDCl_3) δ 12.50 (q, C-18), 17.11 (q, C-16), 17.17 (q, C-19), 20.94 (q, 9- CH_3CO), 21.00 (q, 2- CH_3CO), 21.72 (q, 10- CH_3CO), 26.71 (t, C-7), 31.50 (t, C-6), 35.02 (t, C-14), 37.97 (s, C-15), 40.08 (d, C-3), 44.02 (s, C-8), 45.62 (d, C-1), 70.01 (d, C-10), 71.06 (d, C-2), 74.20 (t, C-17), 76.38 (d, C-5), 76.38 (d, C-9), 97.50 (s, C-13), 114.88 (t, C-20), 130.06 (s, C-11), 142.01 (s, C-12), 147.10 (s, C-4), 170.01 (s, 10- CH_3CO), 170.06 (2- CH_3CO), and 170.06 (s, 9- CH_3CO); FABMS m/z 493 (M+H)⁺; HRFABMS m/z 493.2428 (M+H)⁺, calcd for $\text{C}_{26}\text{H}_{37}\text{O}_9$, 493.2438; ^1H - ^1H COSY correlations (CDCl_3 , H/H): 1/2, 2/3, 5/6, 6/7, 6a/6b, 7a/7b, 9/10, 14a/14b, 17a/17b, and 20a/20b; D-HMBC and HMBC correlations (CDCl_3 H/C): 2/1, 3/1, 16/1, 17a/1, 17b/1, 1/2, 14b/2, 1/3, 7/3, 19/3, 20a/3, 3/4, 6b/5, 20a/5, 5/7, 9/7, 19/7, 2/8, 3/8, 9/8, 19/8, 10/9, 19/9, 9/10, 1/11, 16/11, 17a/11, 18/11, 10/12, 18/12, 1/13, 14a/13, 17a/13, 17b/13, 18/13, 2/14, 10/15, 16/15, 17a/15, 16/17, 3/19, 7a/19, 9/19, 5/20, 2/2-AcO, 9/9-AcO, and 10/10-AcO; NOESY correlations (CDCl_3 H/H): 1/2, 1/17a, 2/9, 2/16, 2/19, 3/7a, 3/10, 3/14b, 3/18, 5/6a, 5/6b, 5/20a, 7a/10, 7a/18, 9/16, 9/19, and 10/18.
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